

# Cardiovascular Effects of Green Tea Catechins: Progress and Promise

Mohammed A. Islam\*

LECOM School of Pharmacy, Bradenton, Florida 34211, USA

Received: May 2, 2012; Accepted: June 3, 2012; Revised: June 2, 2012

**Abstract:** Recently, there is a growing interest in the cardiovascular beneficial effects of green tea. Epidemiological and clinical studies have suggested that consumption of green tea is inversely associated with the risk of developing cardiovascular diseases. Catechins, the major flavonoid constituents of green tea, exert cardioprotective effects through diverse mechanisms that include reversal of endothelial dysfunctions, decreasing inflammatory biomarkers, and providing antioxidant, antiplatelet and antiproliferative effects. Moreover, dietary consumption of green tea catechins has beneficial effects on blood pressure and lipid parameters. This review will focus on discussing the latest research on the cardioprotective effects of green tea catechins and their underlying molecular mechanisms. Several recent patents pertinent to green tea and cardiovascular health will also be discussed. It is noteworthy that clinical studies involving green tea are fraught with multiple complexity and confounding factors. Therefore, a rigorous assessment of the effects of green tea catechins in well-controlled human trials will be required for better understanding of the effects of green tea in cardiovascular health.

**Keywords:** Antioxidant, anti-inflammatory, atherosclerosis, EGCG, endothelial dysfunction, green tea catechins, HDL cholesterol, LDL cholesterol.

## INTRODUCTION

Green tea has been considered as a healthy beverage for centuries in East Asian countries [1]. The consumption of green tea has increased in western countries as a refreshing beverage or as dietary supplements containing dried leaves or extract because of its various alleged health benefits [2, 3]. Consequently, there is an increasing research interest in the effects of green tea in human health [4].

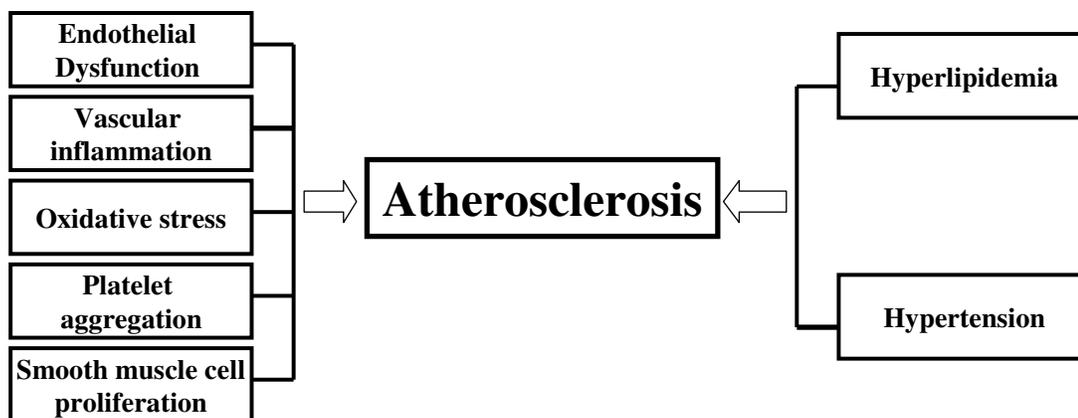
Green tea, similar to black and oolong tea, is produced from the leaves of *Camellia Sinensis*. However, a different processing technique is used that involves quick steaming of the freshly harvested green tea leaves, so that the oxidative enzymes are inactivated and the flavonoid contents are retained during the subsequent rolling and drying processes. The chemical composition of green tea varies due to differences in origin, variety, growing conditions, and production processes [5-7]. The polyphenol contents constitute up to 30% of the dry weight which include flavonoids, flavanols, flavandiols, and phenolic acids [8, 9]. More than 70% of flavonoids in green tea are catechins. (-)-Epigallocatechin-3-gallate (EGCG) is the most active compound biologically and constitutes 65% of the total catechins found in green tea. Other green tea catechins include (+)-catechin, (+)-gallocatechin, and (-)-epicatechin [10, 11]. Besides polyphenols, green tea contains caffeine, theophylline, carbohydrates, lipids, proteins, amino acids, sterols, minerals and trace elements, vitamins (B, C, E), pigments, and some volatile aldehydes and ketones, etc [9]. The over-the-counter

green tea supplements are available as concentrated extracts of green tea containing 200-400 mg of EGCG per dose along with caffeine and other polyphenols.

Cardiovascular disease (CVD) is the leading cause of morbidity and mortality in industrialized countries. According to World Health Organization report, CVDs were accounted for 29% of all global deaths in 2004 and about 23.6 million people will die from CVDs by 2030 [12]. In USA, CVD accounts for 1 of every 3 deaths and more than one-fourth of the U.S. populations live with CVD [13]. Clinical manifestations and pathophysiology of CVD are complex and multifactorial (Fig. (1)). In most instances, CVD is attributed to atherosclerosis, a vascular chronic inflammatory condition.

The endothelium is the intrinsic regulator of vascular tone, local inflammation, and angiogenesis via the secretion of various vasoactive substances [14]. Endothelial dysfunction is considered as an early marker of atherosclerosis. The severity of atherosclerosis predicts cardiovascular events such as myocardial infarction, heart failure, cardiac arrhythmias, peripheral arterial disease, and stroke [15]. The *in vivo* endothelial function is assessed by exploiting several technical approaches such as measurement of flow-mediated vasodilation (FMD), arterial compliance, coronary artery diameter, and coronary blood flow. Moreover, altered plasma levels of cytokines, intracellular cell adhesion molecules (ICAM) [16-18], soluble vascular cell adhesion molecules (VCAM-1), plasma nitric oxide (NO) metabolites [19], and C-reactive protein (CRP) [20], commonly serve as circulating predictors of endothelial dysfunction. Additionally, altered levels of endothelin-1 (ET-1) and Von Willebrand factor (vWF) also serve as predictors of endothelial dysfunction

\*Address correspondence to this author at the LECOM Bradenton School of Pharmacy, 5000 Lakewood Ranch Blvd., Bradenton, FL 34211, USA; Tel: (941) 782-5694; Fax: (941) 782-5724; E-mail: mislam@lecom.edu



**Fig. (1).** Pathophysiology and risk factors associated with atherosclerosis

[21, 22]. The pro-inflammatory cytokines, induction of oxidative enzymes, and uncoupling of mitochondrial electron transport are involved in the generation of reactive oxygen species (ROS) which, in turn, contributes to endothelial dysfunction [23, 24] leading to atherosclerosis and myocardial infarction.

The cardiovascular effects of green tea consumption have been extensively studied in human as well as in animal models [25, 26]. There are accumulating evidences from epidemiological and human interventional studies that consumption of green tea is associated with decreased CVD risks [27-30]. The cardiovascular as well as other potential health benefits of green tea consumption have been previously reviewed [1, 31]. Moreover, several recent reviews have focused on the discussion of the underlying mechanisms of cardioprotective effects of green tea catechins [25, 26, 32, 33]. In humans, most of the studies involved the use of green tea beverages or tea extracts. Therefore, it is uncertain whether the cardiovascular benefits are conferred by flavonoids or by other constituents of green tea such as caffeine or theophylline. In addition, variations in green tea sources, manufacturing process, and flavonoid types and contents may further act as confounding factors in the assessment of cardioprotective effects of green tea catechins. Thus, it is important to explore the cardiovascular effects of pure isolated green tea catechins. However, the studies involving isolated pure green tea flavonoids in humans are limited. The present review summarizes current developments on the cardiovascular effects of green tea catechins with special emphasis on recent studies exploring the underlying molecular mechanisms. In addition, studies involving bioavailability are also discussed.

## CARDIOVASCULAR DISEASES AND GREEN TEA

### Human Studies

Accumulating evidence from population-based studies have suggested an inverse association between consumption of green tea and the risks of CVD [34-38]. Table 1 shows a summary of human interventional studies of green tea and tea catechins in endothelial dysfunction, vascular reactivity and atherosclerosis. Several studies in Japanese population reported significant reduction in mortality from all causes

and CVD in individuals who consumed five or more cups of green tea compared with those who consumed less than one cup daily or none [36, 39, 40]. Green tea was found to be beneficial in patients with angiographically proven coronary artery disease (CAD) [41]. In a recent study in Chinese population, Wang *et al.* [42] showed that green tea consumption was associated with decreased prevalence of coronary artery disease that was defined as having at least one significant coronary artery stenosis. Intriguingly, this reduced risk of CAD was observed in male, but not in female patients. A number of human interventional studies examined the effects of green tea flavonoids on endothelium-dependent vascular reactivity and biomarkers for CVD [43-46]. Altered FMD of the brachial artery is an independent predictor of cardiovascular risk. The effect of green tea on brachial artery reactivity was studied in 14 healthy individuals (age 30+/-3 years) with no cardiovascular risk factors except from smoking (50%). Intake of 6 g of green tea significantly increased FMD (by 3.69%, peak at 30 min), but it did not have any effect on high-sensitivity CRP, IL-6, IL-1 $\beta$ , total plasma antioxidative capacity, or total plasma oxidative status suggesting an acute beneficial effect on endothelial function [46]. In another study, consumption of green tea extract for 5 weeks caused enhanced FMD in healthy women (n=14) compared with placebo [47]. Similarly, green tea consumption significantly improved vascular reactivity, increased circulating endothelial progenitor cells, and reduced oxidative stress biomarker, 8-isoprostaglandin -F<sub>2 $\alpha$</sub>  in chronic smokers. In male smokers, consumption of green tea (8 gm/day) for 2 weeks significantly improved FMD [48]. In contrast to the above studies, a recent study showed that daily consumption of high doses of green tea extract did not significantly alter endothelial-dependent and -independent vascular reactivity [44].

A number of studies assessed the effects of acute and chronic exposures of green tea catechins on endothelial function in human. Loke *et al.* have recently shown that acute per oral treatment with pure dietary flavonoid, (-)-epicatechin (200 mg) improves endothelial function in healthy volunteers by augmenting NO status and reducing ET-1 plasma concentrations [49]. Acute and chronic treatment with high dose (580 mg) green tea catechins significantly increased the forearm blood flow (FBF) response in male smokers.

**Table 1. Human studies: endothelial dysfunction, vascular reactivity and atherosclerosis.**

Study	Isoflavone Treatment	Outcomes	References
Placebo-controlled, parallel; Healthy men (n=31)	Green tea extract (714 mg green tea polyphenols)/day x 3 weeks	No changes in cardiovascular biomarkers	[44]
Healthy women (n=14)	Taken catechol or placebo for 5 weeks (catechol capsules contained caffeine 150 mg, 375 mg of catechins that included 270 mg EGCG)/day	No changes in total cholesterol, HDL or LDL ↓FMD ↓Oxidized LDL ↓Triglycerides	[47]
Randomized, placebo-controlled, cross-over trial; Healthy men (n=12)	(-)-epicatechin 200 mg, (acute oral administration) Control group-water	↑NO status (plasma S-nitrosothiols) ↓ Plasma endothelin-1 level	[49]
Male healthy smokers (n=30) divided in three groups	Green tea beverage: Control group: 0 mg, middle dose group: 80 mg and high dose group: 580 mg of tea catechins x 2 weeks	↑ FBF, ↑ NO ↓Asymmetrical dimethylarginine, malondialdehyde, CRP, MCP-1, and soluble CD40 ligand	[50]
Healthy male non-smokers (n=22)	7 cups of water/day x 2weeks, then 7 cups of green tea/day x2 weeks	↓LDL oxidation in vivo No effects on platelet aggregation, platelet thromboxane production, or plasma MMPs concentration	[57]
Healthy females (n=5)	Green tea 1.5 g/3 times/day x 2 weeks, then wash out x 1 week, then water only x 2 weeks, green tea x 2 weeks	↓Susceptibility of plasma LDL to oxidation, ↓LDL	[58]
Randomized, double-blind, placebo-controlled; Obese women (n=78 with placebo 37)	Green tea extract (400 mg) 3 times/day x 12 weeks	↓ Triglycerides ↓ Lipid peroxidation ↓LDL ↑HDL ↑Adiponectin	[60]
Double-blind, parallel multicenter trial; Japanese men and women (n=240; treatment 123 and control 117)	Green tea (583 mg catechins) and control (96 mg catechins)/day x 12 weeks	↓Systolic BP ↓LDL ↓Body fat	[61]
Randomized, double-blind, placebo-controlled; Healthy postmenopausal women (n=103)	EGCG (400-800 mg) in capsules/day x 2 months	LDL ↓ Beneficial effects on glucose-related markers	[62]
Healthy men and women (n=40, catechin group 29, control 11)	Polyphenon 70 capsules containing 500 mg catechins once a day x 4weeks	No influence on plasma LDL ↓Circulating Ox-LDL	[63]
Healthy volunteers on controlled diet	Treatment group: 2 cups containing 250 mg total catechins with controlled diet x 6 weeks Control group: only controlled diet	↓Induced DNA oxidative damage in lymphocytes ↓Plasma peroxide levels ↓LDL ↑Plasma antioxidant activity	[64]
Randomized, controlled, prospective; Adults with obesity and metabolic syndrome (n=35)	Green tea beverages (4 cups/day) or extract (2 capsules/day) x 8 weeks	↓ Lipid peroxidation ↓LDL	[65]
Randomized, double-blind, cross-over setting; Healthy endurance-trained men (n=10)	Green tea extract (containing 160 mg total catechins where 70 mg was EGCG/day) x 3 weeks	No effects on lipid parameters, inflammation processes, and oxidative stress.	[66]

(Table 1) Contd....

Study	Isoflavone Treatment	Outcomes	References
Randomized, double-blind, placebo-controlled; Mild to moderately hypercholesterolemic subjects (n=102, men 67 and women 35)	Theaflavins (77.5 mg) or Theaflavins/Catechins (75 and 150 mg, respectively) plus 195 mg other polyphenols in capsules/day	No effects on LDL, triglycerides	[67]
Randomized, controlled; Overweight or obese males (n=46, placebo n=42)	EGCG capsules 400 mg, twice daily x 8 weeks	↓Diastolic blood pressure No effect on insulin sensitivity	[72]
Randomized, controlled, cross-over trial; Males (40-69 yrs)	Decaffeinated green tea extract (400 mg total catechins)/capsules/twice daily x 6 weeks	No effects on BP or biomarkers of metabolic function	[73]

Moreover, chronic consumption of high-dose catechins caused a significant increase in plasma NO associated with decreased levels in malondialdehyde, 4-hydroxynonenal, CRP, monocyte chemotactic protein-1 (MCP-1), and soluble CD40 ligand, and dimethylarginine [50]. In patients with coronary artery disease, daily consumption of green tea or tea flavonoids improved endothelial function [51-53].

Dyslipidemias resulting from elevated levels of LDL-cholesterol, phospholipids, triglycerides, and low plasma HDL-cholesterol levels act as a predisposing factor for the development of atherosclerotic plaques [54]. In a cross-sectional study among 1371 Japanese men, Imai *et al.* [55] found that increased green tea consumption was associated with decreased plasma levels of LDL, total cholesterol, and triglycerides, but increased levels of HDL. After meta-analysis of 14 eligible randomized controlled trials with a population of 1136, Zheng *et al.* [56] concluded that consumption of green tea catechins is associated with significant reduction in total and LDL cholesterol levels, but not in HDL cholesterol. Green tea-mediated decreased plasma levels of total cholesterol and LDL cholesterol were consistently reported in a number of studies, however, the findings on the effects on triglycerides and HDL-cholesterol were mixed and inconsistent [57-65]. Moreover, several recent randomized, placebo-controlled trials involving healthy or moderately hypercholesterolemic subjects reported no effects of green tea catechins on lipid parameters [47, 66, 67].

Hypertension is the most common form of CVD and is one of the major risk factors for cardiovascular-related mortality. Green tea consumption has been believed to exert antihypertensive effects. Epidemiological evidence regarding the beneficial effects of green tea on blood pressure is inconsistent [68, 69]. In Chinese population, habitual consumption of 120 mL /day or more of green tea was reported to reduce the risk of developing hypertension [70]. Similarly, in another study in older women, Hodgson *et al.* reported that long-term consumption of green tea may lower the risk of hypertension [71]. In a double-blind, parallel multicenter trial in Japan, green tea catechins have shown to decrease systolic blood pressure [61]. Similarly, reduction in diastolic blood pressure was observed in obese males who took EGCG (800 mg/day in divided doses for 8 weeks) [72]. However, oral administration similar doses of decaffeinated EGCG for 6 weeks failed to alter blood pressure [73].

### Animal Studies

A number of animal models have been exploited to explore the cardiovascular effects of green tea catechins and their underlying molecular mechanisms. As summarized in Table 2, green tea catechins have been reported to impart cardioprotective effects through improvement of vascular reactivity, reduction of inflammation, potent antioxidant activity, and prevention of platelet aggregation and vascular smooth muscle cell (VSMC) proliferation. Moreover, green tea catechins were found to reduce the risk of cardiovascular diseases through improved cholesterol metabolism and anti-hypertensive effects. Interaction of green tea catechins with a number of molecular targets and cell signaling pathways underlie their cardiovascular beneficial effects.

### EFFECTS OF GREEN TEA ON VASCULAR HOMEOSTASIS

There have been extensive *in vitro* as well as *in vivo* animal studies to test the effects of green tea flavonoids in vascular homeostasis. Acute treatment of isolated rat thoracic aortas with green tea catechins resulted dose-dependent vasodilation against phenylephrine-induced contractions [74-76]. Similarly, in rat mesenteric veins, EGCG (1-100  $\mu$ M) demonstrated a dose-dependent vasodilation which was blocked by L-NAME (a NO synthase agonist) or the phosphatidylinositol-3-kinase (PI3K) inhibitor [74].

The effects of green tea catechins against vascular dysfunction were evaluated in different animal models of chronic cardiovascular and metabolic disorders. In spontaneously hypertensive rats, daily supplementation of 200 mg EGCG/kg for 3 weeks significantly improved vascular tone of mesenteric vascular beds and reduced systolic blood pressure [74]. Ihm *et al.* [77] found that daily treatment with 30 mg/kg catechin for 12 weeks significantly reduced blood pressure, fasting sugar, and improved endothelium-dependent aortic relaxation in an *ex vivo* setting in prediabetic Otsuka Long-Evans Tokushima Fatty (OLETF) rats. Thus, green tea flavonoids may confer beneficial effects against endothelial dysfunction and hyperglycemia. Consistently, in Goto-Kakizaki (GK) rats with type 2 diabetes, the green tea catechins improved blood pressure and blood glucose levels [78]. Of interest, in a rat model of angiotensin-II-

**Table 2. Mechanisms by which green tea catechins confer cardioprotection.**

Reversal of Endothelial Dysfunction:
<ul style="list-style-type: none"> <li>• ↑NO</li> <li>• ↑eNOS activity</li> <li>• ↑cGMP</li> <li>• ↑PI3K/Akt</li> <li>• ↓ET-1 expression</li> </ul>
Reduction of Vascular inflammation:
<ul style="list-style-type: none"> <li>• ↓VCAM-1</li> <li>• ↓Cytokines</li> <li>• ↓ Chemokines</li> <li>• ↓ Adhesion molecules</li> <li>• ↓MCP-1 mRNA and protein expression</li> <li>• ↓Leucocyte infiltration</li> <li>• ↓p38 MAPK and NF-kappa B activation</li> </ul>
Antioxidant effects:
<ul style="list-style-type: none"> <li>• ↑Antioxidant enzymes (superoxide dismutase, catalase, and glutathione peroxidase)</li> <li>• ↓Pro-oxidant enzymes</li> <li>• Scavenging free radicals</li> <li>• Chelating redox ions</li> <li>• ↓OxLDL</li> <li>• ↓LOX-1 mediated signaling</li> </ul>
Antiplatelet and antithrombotic activity:
<ul style="list-style-type: none"> <li>• ↓Intracellular Ca<sup>2+</sup> mobilization</li> <li>• ↓Inositol 1,4,5-triphosphate formation</li> <li>• ↓Binding of fibrinogen-GPIIb/IIIa</li> <li>• ↓Phospholipase C<sub>γ</sub> phosphorylation</li> <li>• ↓Protein tyrosine phosphorylation</li> <li>• ↓Arachidonic acid liberation</li> <li>• ↑Cellular prostaglandin D<sub>2</sub> levels</li> </ul>
Inhibition of cardiomyocyte apoptosis during oxidative myocardial injury:
<ul style="list-style-type: none"> <li>• ↓Expression of telomere repeat-binding factor 2 (TRF-2) proteins</li> <li>• ↑p53 and p21 protein levels</li> </ul>
Inhibition of VSMC Proliferation:
<ul style="list-style-type: none"> <li>• Arrest VSMCs in G<sub>1</sub> phase by inactivating pAkt</li> <li>• ↓Expression of VEGF by blocking PDGF and ERK1/2 signaling</li> <li>• ↓Matrix protein degradation</li> <li>• ↓Cell migration through ↓expression of MMP-2 and MMP-9</li> </ul>

induced hypertrophy, treatment with green tea extract prevented hypertension and target organ damage [79, 80]. A mixture of dietary supplements containing green tea catechins among other constituents strongly reduced the development of atherosclerotic lesion in female ApoE\*3Leiden

transgenic mice [81]. Similar effects were observed in atherosclerosis susceptible C57BL/6J, apolipoprotein E-deficient mice [82]. In Apolipoprotein E-null mice, administration of EGCG (10 mg/kg, i.p., 42 days) significantly reduced carotid artery injury-induced atherosclerotic plaque formation; however, EGCG failed to reduce the established atherosclerotic plaque [83]. Similarly, in LDL receptor knockout mice, green tea catechins when given with high cholesterol diet prevented the development of atherosclerosis and target organ damage when compared with control group.

## ENDOTHELIAL FUNCTION AND NO METABOLISM

The endothelium maintains the vascular homeostasis through a balanced regulation of vasodilation and vasoconstriction [84]. NO, synthesized from arginine by endothelial nitric oxide synthase (eNOS), acts as a vasodilator as well as prevents leukocytes adhesion, platelet aggregation, and release of pro-inflammatory cytokines [85]. In an impaired endothelium, induction of pro-inflammatory endothelial phenotype and reduced generation of NO promote progression of atherosclerosis and future cardiac events [84, 86, 87]. There is convincing evidence from animal studies that green tea catechins exert beneficial effects against endothelial dysfunction by increasing NO production. Recent studies provided evidence that EGCG resulted in endothelium- and NO-dependent vasodilation in rat aortic rings [88, 89]. Benito *et al.* further showed that the endothelium-dependent vasodilation involved increased eNOS activity, NO production, and cyclic guanosine-3',5'-monophosphate (cGMP) production [90]. Consistently, green tea catechins concentration-dependently increased NO production in human umbilical cord endothelial cells (HUVEC) [91] and bovine aortic endothelial cells [88]. In consistent with *in vitro* animal studies, studies in cultured endothelial cells have also documented that the EGCG-induced increased NO production involves increased activity of eNOS, phosphatidylinositol-3-hydroxy kinase, and Akt signaling pathways [74, 92].

NO and ET-1 interplay as natural counterparts in maintaining the vascular tone. Recent data suggest that an imbalance between these two mediators contributes to endothelial dysfunction leading to the progression of cardiovascular diseases [93]. ET-1 is a potent vasoconstrictor produced by vascular endothelial cells [94]. Several lines of evidence from human and animal studies suggest that elevated levels of ET-1 is implicated in endothelial dysfunction leading to abnormal control of vascular tone and blood pressure [95-98]. Recently, Reiter *et al.* [99] have shown that in human aortic endothelial cells EGCG decreased ET-1 expression in endothelial cells via Akt- and AMPK-stimulated FOXO1 regulation of the ET-1 promoter.

## ANTI-INFLAMMATORY EFFECTS

The vascular inflammation is now increasingly considered as a key process in the pathogenesis of atherosclerosis which is initiated with the adhesion of monocytes to the vascular endothelial cells and their subsequent transmigration into sites of inflammation [100]. This atherosclerotic process is dependent on the expression of MCP-1 and interleukin-8 (IL-8) along with several adhesion molecules such as ICAM-1, and VCAM-1, and endothelial leukocyte adhesion mole-

cule-1 (E-selectin) [101, 102]]. Besides these, NO, cytokines, prostacyclin, and cellular processes such as cell growth and differentiation are involved in vascular inflammatory processes. Ramesh *et al.* [103] recently showed that daily administration of EGCG (100 mg/kg, i.p.) significantly decreased the expression of CRP, with concomitant decrease in hematological parameters of inflammation in atherosclerotic rats. Similarly, *in vitro* experiments in VSMCs showed that EGCG concentration-dependently inhibited ET-1-stimulated expression of CRP both in protein and mRNA levels [104].

Inducible nitric oxide synthase (iNOS) contributes to the pathophysiology of atherosclerosis through the generation of NO [105]. Recently, Ahn *et al.* [106] reported increased expression of iNOS in HUVEC. Moreover, in HUVEC, EGCG and ECG dose-dependently inhibited cytokine-induced VCAM-1 expression and monocyte adhesion [107]. In LDL receptor knock-out mice, catechin administration significantly suppressed VCAM-1 expression in atherosclerotic lesions [108]. Green tea catechins have been found to reduce the expression of pro-inflammatory cytokines, chemokines, and adhesion molecules that are regulated by the nuclear transcription factor NF-kappa B [109, 110]. Moreover, Hong *et al.* reported that tea catechins inhibited phorbol 12-myristate 13-acetate-induced MCP-1 mRNA and protein expression in human endothelial cells and thereby reduced leucocyte infiltration and their subsequent transmigration through cultured human endothelial monolayers by suppressing p38 MAPK and NF-kappa B activation [111]. In consistent with the above studies, EGCG reduced TNF- $\alpha$ -induced MCP-1 production in bovine coronary artery endothelial cells by inhibiting Akt phosphorylation as well as TNF activation of TNFR1 [112]. EGCG suppresses pro-inflammatory actions of TNF- $\alpha$  through over expression of heme oxygenase-1 in vascular endothelial cells via p38 MAPK and Nrf-2 signaling pathways [113].

## ANTIOXIDANT EFFECTS

There is a growing number of evidence that green tea catechins exert antioxidant activity through a number of mechanisms that include induction of antioxidant enzymes, inhibition of pro-oxidant enzymes, scavenging free radicals, chelating redox ions, and inhibiting transcription factors for redox reaction [114]. In animal models, green tea catechins have been reported to upregulate antioxidant enzymes such as superoxide dismutase, catalase and glutathione peroxidase [115, 116]. In apolipoprotein E null mice, a mouse model of human atherosclerosis, EGCG increases the antioxidant capacity in systemic circulation as well as in local vascular tissues [83]. Catechins and their metabolites, (-)-epicatechin-5-O- $\beta$ -glucuronide and (+)-catechin-5-O- $\beta$ -glucuronide, serve as electron donors and highly efficient scavengers of free radicals such as nitric oxide (NO), peroxynitrite, superoxide anions, and singlet oxygen [114, 117].

The cellular proliferative and apoptotic processes involve catalytic generation of superoxide free radicals through endothelial NADPH oxidase, a multisubunit enzymatic complex comprising of two membranes bound subunits, gp91 and p22<sup>phox</sup>. The assembly of the active NADPH complex is regulated by two cytoplasmic subunits (p47<sup>phox</sup> and p67<sup>phox</sup>) and a G-protein, Rac-1 [118]. In vascular endothelial cells,

activation of Rac-1 and p47<sup>phox</sup> is involved in the generation of superoxide that stimulates inflammatory gene expression through redox-sensitive signaling mechanisms [119]. Green tea catechins have been reported to inhibit NADPH oxidase activity leading to the reduced production of ROS [120]. In a recent study in the prediabetic OLETF rat model, daily catechin treatment (30 mg/kg for 12 weeks) reduced the expression of p22<sup>phox</sup> and p47<sup>phox</sup> NADPH oxidase subunits as well as ROS formation in aortic endothelial cells [77].

Oxidative modification of LDL (ox-LDL) plays an important role in the development of atherosclerosis. High level of ox-LDL triggers inflammatory reactions, decreases activities of antioxidant enzymes such as superoxide dismutase and glutathione peroxidase, and causes lipid peroxidation leading to endothelial dysfunction and formation of atherosclerotic plaques [121]. Earlier studies reported that catechins can inhibit the oxidation of LDL both *in vitro* and in animal studies [58, 122-125]. In vascular endothelial cells, activation of LOX-1, a lectin-like receptor for oxLDL, has been suggested to increase intracellular ROS, including H<sub>2</sub>O<sub>2</sub> and singlet oxygen [126]. Recently, Ou *et al.* have shown that EGCG exerted significant cytoprotective effects in cultured human umbilical vein endothelial cells through inhibition of the oxLDL-induced LOX-1 mediated signaling pathway [127].

During cardiac dysfunction involving oxidative myocardial injury, telomere signalling plays a role in regulating cardiomyocyte apoptosis through decreased expression of telomere repeat-binding factor 2 (TRF-2) proteins and upregulation of p53 and p21 protein levels. *In vitro* experiments in H9c2 cells have demonstrated that EGCG suppresses H<sub>2</sub>O<sub>2</sub>-induced cardiomyocyte apoptosis and telomere attrition [128].

## ANTI-PROLIFERATIVE EFFECTS

During atherosclerosis, the intimal and medial thickening contributes to vascular VSMC proliferation and migration. In animal models, EGCG has been shown to suppress of VSMC proliferation induced by high glucose [129], angiotensin-II [130], and platelet derived growth factor (PDGF) [131, 132]. EGCG treatment has been shown to arrest VSMCs in G<sub>1</sub> phase of the cell cycle by inactivation of pAkt, leading to subsequent apoptosis [131]. Similarly, catechins have been found to inhibit angiotensin-II-induced VSMC proliferation in rat aorta through inhibition of mitogen-activated PK signaling [130]. Animal studies and cell culture experiments revealed that the anti-proliferative effects of green tea catechins were mediated through the inhibition of a number of signaling pathways including mitogen-activated PK, downstream PI3K/Akt PKC, and ERK1/2 signaling [131, 132]. In human VSMCs, EGCG has been reported to dose-dependently decrease the expression of vascular endothelial growth factor (VEGF) expression via blocking PDGF and ERK1/2 signaling [133]. A further study showed that EGCG inhibited cell proliferation by blocking the interaction of PDGF with its receptor [134].

VSMCs migrate from the tunica media to the sub-endothelial region during atherogenesis and in response to vascular injury that may occur clinically after organ transplantation, stent implantation, or angioplasty. Matrix metal-

loproteinases (MMPs), particularly, MMP-2 and MMP-9 have been implicated in cell migration and tissue remodeling process due to their ability of degrading extracellular matrix proteins [100, 135]. Catechins, in a dose-dependent manner, have been shown to prevent matrix protein degradation and cell migration by inhibiting expression or activity of MMP-2 [136, 137] and MMP-9 [138]. Gouni-Berthold *et al.* showed that catechins exert their antiproliferative and apoptotic effects through the inhibition of growth factor-RTK-mediated signal transduction pathways. Enhanced RTK activity has been associated with the development of atherosclerosis [139]. Thus, above *in vitro* studies suggest a potential anti-atherogenic action of catechins at pharmacological doses. However, the relevance of these *in vitro* findings to clinical situation remains to be further evaluated.

### ANTIPLATELET AND ANTITHROMBOTIC ACTIVITY

Platelets play an important role in the development of acute thrombotic events as well as in the pathogenesis of atherosclerosis. Vascular endothelial injury promotes rapid platelet hyper-reactivity and aggregation leading to a formation of thrombi which could trigger acute myocardial infarction [140]. *In vitro* and *in vivo* studies have reported potential antiplatelets and antithrombotic effects of tea catechins [141-144]. Catechins dose-dependently inhibit various stimuli-induced *in vitro* platelet aggregation of humans [145, 146] and animals [143, 144]. Exposure of human platelet concentrates to EGCG (50 and 100 microgram/ml) maintained platelet aggregability and preserved the platelets by inhibiting the activation and apoptosis [145]. There is accumulating evidence that tea catechin supplementation improved platelet aggregation and thrombosis through interacting with a diverse signaling mechanisms [147]. In human platelets, catechins inhibit intracellular calcium mobilization *via* activation of  $Ca^{2+}$ -ATPase and inhibition of inositol 1,4,5-triphosphate formation leading to the attenuation of fibrinogen-GPIIb/IIIa binding [142]. Recently, Jin *et al.* [144] have shown that antiplatelet activity of EGCG is attributable to interaction with multiple cellular targets such as inhibitions of phospholipase  $C\gamma_2$  phosphorylation, protein tyrosine phosphorylation, arachidonic acid liberation, and elevation of cellular prostaglandin  $D_2$  levels.

Inflammatory conditions are associated with increased release of platelet activators such as arachidonic acid, prostaglandins, endoperoxides, and thromboxane  $A_2$  [148]. Sugatani *et al.* [149] have shown that tea catechins reduced PAF- and TPA-induced rabbit platelet aggregation through inhibiting acetyl-CoA:1-alkyl-sn-glycero-3-phosphocholine acetyltransferase enzyme. Several other studies have explored the antiplatelet and antithrombotic effects of tea catechins through their suppressing effects on arachidonic acid release and thromboxane  $A_2$  synthase activity [143, 144].

### EFFECTS ON LIPID METABOLISM

Tea catechins have been reported to favorably affect lipid metabolism in a number of animal models. In rats with hypercholesterolemia, EGCG improves hyperlipidemia by reducing LDL-cholesterol, but elevates HDL-cholesterol levels [150-153]. Similarly, improved blood glucose and lipid pro-

files have been observed in diabetic rats that received EGCG (25 mg/kg/day) for 8 weeks [154]. Studies in animal models explored several possible mechanisms explaining the effects of green tea catechins on lipid metabolism. Green tea catechins modulate cholesterol metabolism mainly by targeting its biosynthesis [155] and intestinal absorption [156-158]. Hydroxy-3-methyl-glutaryl-CoA reductase (HMGR) is the rate-controlling enzyme in the biosynthesis of cholesterol. Cuccioloni *et al.* [159] have shown that EGCG potently inhibits the *in vitro* activity of HMGR by competitive binding to the cofactor site of the enzyme. In addition, catechins may also have direct inhibitory effect on squalene epoxidase, another rate limiting enzyme of cholesterol biosynthesis [160]. Catechins enhance intestinal lipid absorption through direct interference with the emulsification, hydrolysis, micellar solubilization of lipids, formation of insoluble co-precipitates of cholesterol, and interaction with pancreatic phospholipase  $A_2$  [156, 157, 161-163]. Moreover, tea catechins have also been reported to modulate specific transport proteins located on the brush border membranes that play a significant role in lipid uptake by enterocytes [164]. Green tea catechins enhances cholesterol 7  $\alpha$ -hydroxylase mRNA expression and promoter activity in human hepatoma cells which may underlie the hypocholesterolemic effects of catechins [165]. Using DNA microarray analysis and real-time PCR technique, Goto *et al.* [166] recently demonstrated that EGCG treatment improved cholesterol metabolism by up-regulating LDL receptor mRNA expression level and decreasing extracellular apoB levels.

### CURRENT & FUTURE DEVELOPMENTS

Extensive studies have been performed on the cardioprotective effects of green tea and tea catechins in human as well as *in vitro* and *in vivo* animal models. The health benefits of green tea catechins such as maintenance of endothelial function, vascular homeostasis, and associated reduction of CVD risks are becoming increasingly recognized. There are mounting evidences emerged from epidemiological [28-30, 36], human interventional [46, 47, 50, 59] as well as animal studies [74-78, 80].

Several recent patents have described the promising effects of green tea catechins against cardiovascular diseases. Epicatechins and derivatives and their salts have been found to reduce of infarct size in the heart with total coronary occlusion in animals as well as in humans [167]. Bukowski and Walters patented their intriguing discovery of cardiovascular beneficial effects of tea-derived compositions comprising predetermined amounts of EGCG and L-theanine. They found that encapsulated tea components of defined composition when given orally twice daily for 12 weeks significantly lowered blood pressure, LDL cholesterol, and chronic inflammation as measured by decreased serum amyloid alpha levels [168]. Several other recent patents described the use of green tea extracts or tea catechins as an ingredient of pharmaceutical compositions intended for the treatment of cardiovascular and other ailments [169-171].

There are substantial inconsistencies in the published data emerged from human interventional studies that investigated the effects of green tea or its flavonoids on CVD markers [36, 44]. In a recent randomized controlled trial,

consumption of catechin-enriched GT (100- 400 mg/day) for 9 weeks did not affect serum adiponectin, one key factor in atherosclerosis, and other cardiovascular disease (CVD) risk factors in apparently healthy subjects [172]. As previously discussed, green tea exerted antiatherosclerotic benefits in patients with angiographically proven coronary artery disease [41], however, Hirano *et al.* [173] reported no relationship between green tea consumption and coronary artery disease in 393 patients who underwent coronary angiography. Moreover, in a meta-analysis of 30 randomized control trials, seventeen studies provided evidence of significant beneficial effects of green tea against CVD [36]. It is difficult to explain these discrepancies because most of the published studies related to cardioprotective effects of green tea and tea catechins in humans suffer from limitations of poor study design, short study duration, and small sample size. Moreover, the evaluation of cardioprotective effects of green tea and tea catechins in humans is difficult because of other confounding factors such as different forms of green tea supplements, variable amounts of catechins, their interactions with other potentially active ingredients such as caffeine, and food matrix affecting the bioavailability of green tea catechins. The bioavailability of the dietary nutrient is an important factor in the determination of the beneficial effect. It is possible that the bioavailability of one key catechin can be affected by the interactions with other competitive nutrients in the level of absorption, metabolism, and membrane transportation. After ingestion, green tea catechins are absorbed and metabolized and appear in blood and urine [174, 175]. However, little is known about how catechins absorption is affected by food matrix effect. In a recent study, Auger *et al.* [176] demonstrated that co-ingestion of green tea catechins with bread, cheese, or glucose did not significantly affect the absorption, metabolism, and excretion of tea catechins. However, Chow *et al.* [177] reported that bioavailability and the pharmacokinetic profile of EGCG is enhanced when taken orally in the fasting condition. Enhance the bioavailability of EGCG has been reported when administered as encapsulated nanolipid particles in comparing with free EGCG [178-180]. Improved bioavailability of green tea catechins have been reported from formulations stabilized with addition of natural antioxidants such as vitamin C or vitamin C analogs [181]. Similarly, modification of green tea polyphenol formulations by adding one or more ester-linked fatty acids have also been shown to enhance bioavailability [182].

Animal and cell line studies with isolated green tea flavonoids and catechins have shown promising cardioprotective effects and mechanistic insights on their effects. Green tea catechins exert cardiovascular effects through multiple mechanisms, including, improved vascular homeostasis through NO signaling, anti-inflammatory, antioxidant, anti-proliferative, and anti-thrombogenic mechanisms. Moreover, in ischemic myocardium, EGCG has been reported to inhibit apoptotic cell death by inhibiting STAT-1 activity [183]. EGCG has also been found to suppress oxidative stress-induced apoptosis and telomere attrition in cardiac cells [128]. However, extrapolation of the results of animal or cell culture studies to human clinical situations is uncertain mainly because of the concentrations of green tea catechins used in animal or *in vitro* studies (usually 1  $\mu\text{M/L}$  to 100

$\mu\text{M/L}$ ) are far higher than the plasma concentrations achievable in human after green tea consumption [184]. It is difficult to achieve therapeutic concentration ( $\sim 10 \mu\text{M/L}$ ) by ingesting high dose (800 mg capsules of pure EGCG by mouth [185]. Moreover, the results from animal studies should be interpreted with caution due to the inherent biological variations as well as pharmacokinetic and pharmacodynamic differences of the test agents in animals versus humans. Although current evidences of the cardioprotective effects of green tea catechins from human as well as animal studies are promising, further rigorous and well-designed human interventional trials are warranted to provide better understanding of the effects of green tea in cardiovascular health.

## ACKNOWLEDGEMENTS

The author thanks Drs. Deepak Gupta and Chandan Thomas for critically reading the manuscript.

## CONFLICT OF INTEREST

There is no conflict of interest.

## REFERENCES

- [1] McKay DL, Blumberg JB. The role of tea in human health: An update. *J Am Coll Nutr* 2002; 21: 1-13.
- [2] Sato T, Miyata G. The nutraceutical benefit, part I: Green tea. *Nutrition* 2000; 16: 315-7.
- [3] Mak JC. Potential role of green tea catechins in various disease therapies: Progress and promise. *Clin Exp Pharmacol Physiol* 2012; 39: 265-73.
- [4] Erdman JW, Jr., Balentine D, Arab L, Beecher G, Dwyer JT, Folts J, *et al.* Flavonoids and heart health: proceedings of the ILSI North America Flavonoids Workshop, May 31-June 1, 2005, Washington, DC. *J Nutr* 2007; 137: 718S-37S.
- [5] Khokhar S, Magnusdottir SG. Total phenol, catechin, and caffeine contents of teas commonly consumed in the United Kingdom. *J Agric Food Chem* 2002; 50: 565-70.
- [6] Lin YS, Tsai YJ, Tsay JS, Lin JK. Factors affecting the levels of tea polyphenols and caffeine in tea leaves. *J Agric Food Chem* 2003; 51: 1864-73.
- [7] Sabhapondit S, Karak T, Bhuyan LP, Goswami BC, Hazarika M. Diversity of catechin in northeast Indian tea cultivars. *Scientific World Journal*; 2012: 485193.
- [8] Graham HN. Green tea composition, consumption, and polyphenol chemistry. *Prev Med* 1992; 21: 334-50.
- [9] Chacko SM, Thambi PT, Kuttan R, Nishigaki I. Beneficial effects of green tea: a literature review. *Chin Med* 2010; 2010; 5: 13.
- [10] Sano M, Tabata M, Suzuki M, Degawa M, Miyase T, Maeda-Yamamoto M. Simultaneous determination of twelve tea catechins by high-performance liquid chromatography with electrochemical detection. *Analyst* 2001; 126: 816-20.
- [11] Balentine DA, Wiseman SA, Bouwens LC. The chemistry of tea flavonoids. *Crit Rev Food Sci Nutr* 1997; 37: 693-704.
- [12] WHO. Global atlas on cardiovascular disease prevention and control. Editors: WHO; World Heart Federation; World Stroke Organization 2011.
- [13] Roger VL, Go AS, Lloyd-Jones DM, Benjamin EJ, Berry JD, Borden WB, *et al.* Heart disease and stroke statistics--2012 update: a report from the American Heart Association. *Circulation* 2012; 125: e2-e220.
- [14] Vita JA, Keane JF, Jr. Endothelial function: a barometer for cardiovascular risk? *Circulation* 2002; 106: 640-2.
- [15] Rocha VZ, Libby P. Obesity, inflammation, and atherosclerosis. *Nat Rev Cardiol* 2009; 6: 399-409.
- [16] Hwang SJ, Ballantyne CM, Sharrett AR, Smith LC, Davis CE, Gotto AM, Jr., *et al.* Circulating adhesion molecules VCAM-1, ICAM-1, and E-selectin in carotid atherosclerosis and incident coronary heart disease cases: the Atherosclerosis Risk In Communities (ARIC) study. *Circulation* 1997; 96: 4219-25.

- [17] Wojakowski W, Gminski J. Soluble ICAM-1, VCAM-1 and E-selectin in children from families with high risk of atherosclerosis. *Int J Mol Med* 2001; 7: 181-5.
- [18] Brevetti G, Martone VD, de Cristofaro T, Corrado S, Silvestro A, Di Donato AM, *et al.* High levels of adhesion molecules are associated with impaired endothelium-dependent vasodilation in patients with peripheral arterial disease. *Thromb Haemost* 2001; 85: 63-6.
- [19] Fichtlscherer S, Breuer S, Schachinger V, Dimmeler S, Zeiher AM. C-reactive protein levels determine systemic nitric oxide bioavailability in patients with coronary artery disease. *Eur Heart J* 2004; 25: 1412-8.
- [20] Fichtlscherer S, Rosenberger G, Walter DH, Breuer S, Dimmeler S, Zeiher AM. Elevated C-reactive protein levels and impaired endothelial vasoreactivity in patients with coronary artery disease. *Circulation* 2000; 102: 1000-6.
- [21] Omland T, Lie RT, Aakvaag A, Aarsland T, Dickstein K. Plasma endothelin determination as a prognostic indicator of 1-year mortality after acute myocardial infarction. *Circulation* 1994; 89: 1573-9.
- [22] Blann AD. Is raised von Willebrand factor a marker of endothelial cell damage? *Med Hypotheses* 1993; 41: 419-24.
- [23] Kanani PM, Sinkey CA, Browning RL, Allaman M, Knapp HR, Haynes WG. Role of oxidant stress in endothelial dysfunction produced by experimental hyperhomocyst(e)inemia in humans. *Circulation* 1999; 100: 1161-8.
- [24] Egashira K. Clinical importance of endothelial function in arteriosclerosis and ischemic heart disease. *Circ J* 2002; 66: 529-33.
- [25] Babu PV, Liu D. Green tea catechins and cardiovascular health: an update. *Curr Med Chem* 2008; 15: 1840-50.
- [26] Wolfram S. Effects of green tea and EGCG on cardiovascular and metabolic health. *J Am Coll Nutr* 2007; 26: 373S-88S.
- [27] Peters U, Poole C, Arab L. Does tea affect cardiovascular disease? A meta-analysis. *Am J Epidemiol* 2001; 154: 495-503.
- [28] Keli SO, Hertog MG, Feskens EJ, Kromhout D. Dietary flavonoids, antioxidant vitamins, and incidence of stroke: the Zutphen study. *Arch Intern Med* 1996; 156: 637-42.
- [29] Arts IC, Hollman PC, Feskens EJ, Bueno de Mesquita HB, Kromhout D. Catechin intake might explain the inverse relation between tea consumption and ischemic heart disease: the Zutphen Elderly Study. *Am J Clin Nutr* 2001; 74: 227-32.
- [30] Kuriyama S, Shimazu T, Ohmori K, Kikuchi N, Nakaya N, Nishino Y, *et al.* Green tea consumption and mortality due to cardiovascular disease, cancer, and all causes in Japan: the Ohsaki study. *JAMA* 2006; 296: 1255-65.
- [31] Cabrera C, Artacho R, Gimenez R. Beneficial effects of green tea—a review. *J Am Coll Nutr* 2006; 25: 79-99.
- [32] Stangl V, Dreger H, Stangl K, Lorenz M. Molecular targets of tea polyphenols in the cardiovascular system. *Cardiovasc Res* 2007; 73: 348-58.
- [33] Suzuki J, Isobe M, Morishita R, Nagai R. Tea polyphenols regulate key mediators on inflammatory cardiovascular diseases. *Mediators Inflamm* 2009; 2009: 494928.
- [34] Tokunaga S, White IR, Frost C, Tanaka K, Kono S, Tokudome S, *et al.* Green tea consumption and serum lipids and lipoproteins in a population of healthy workers in Japan. *Ann Epidemiol* 2002; 12: 157-65.
- [35] Hollman PC, Feskens EJ, Katan MB. Tea flavonols in cardiovascular disease and cancer epidemiology. *Proc Soc Exp Biol Med* 1999; 220: 198-202.
- [36] Kuriyama S. The relation between green tea consumption and cardiovascular disease as evidenced by epidemiological studies. *J Nutr* 2008; 138: 1548S-53S.
- [37] Kuriyama S. Green tea consumption and prevention of coronary artery disease. *Circ J* 2010; 74: 248-9.
- [38] Sasazuki S, Kodama H, Yoshimasu K, Liu Y, Washio M, Tanaka K, *et al.* Relation between green tea consumption and the severity of coronary atherosclerosis among Japanese men and women. *Ann Epidemiol* 2000; 10: 401-8.
- [39] Suzuki E, Yorifuji T, Takao S, Komatsu H, Sugiyama M, Ohta T, *et al.* Green tea consumption and mortality among Japanese elderly people: the prospective Shizuoka elderly cohort. *Ann Epidemiol* 2009; 19: 732-9.
- [40] Nakachi K, Matsuyama S, Miyake S, Suganuma M, Imai K. Preventive effects of drinking green tea on cancer and cardiovascular disease: epidemiological evidence for multiple targeting prevention. *Biofactors* 2000; 13: 49-54.
- [41] Sano J, Inami S, Seimiya K, Ohba T, Sakai S, Takano T, *et al.* Effects of green tea intake on the development of coronary artery disease. *Circ J* 2004; 68: 665-70.
- [42] Wang QM, Gong QY, Yan JJ, Zhu J, Tang JJ, Wang MW, *et al.* Association between green tea intake and coronary artery disease in a Chinese population. *Circ J* 2010; 74: 294-300.
- [43] Miller RJ, Jackson KG, Dadd T, Mayes AE, Brown AL, Minihane AM. The impact of the catechol-O-methyltransferase genotype on the acute responsiveness of vascular reactivity to a green tea extract. *Br J Nutr*; 105: 1138-44.
- [44] Frank J, George TW, Lodge JK, Rodriguez-Mateos AM, Spencer JP, Minihane AM, *et al.* Daily consumption of an aqueous green tea extract supplement does not impair liver function or alter cardiovascular disease risk biomarkers in healthy men. *J Nutr* 2009; 139: 58-62.
- [45] Grassi D, Aggio A, Onori L, Croce G, Tiberti S, Ferri C, *et al.* Tea, flavonoids, and nitric oxide-mediated vascular reactivity. *J Nutr* 2008; 138: 1554S-60S.
- [46] Alexopoulos N, Vlachopoulos C, Aznaouridis K, Baou K, Vasiliadou C, Pietri P, *et al.* The acute effect of green tea consumption on endothelial function in healthy individuals. *Eur J Cardiovasc Prev Rehabil* 2008; 15: 300-5.
- [47] Tinahones FJ, Rubio MA, Garrido-Sanchez L, Ruiz C, Gordillo E, Cabrerizo L, *et al.* Green tea reduces LDL oxidability and improves vascular function. *J Am Coll Nutr* 2008; 27: 209-13.
- [48] Kim W, Jeong MH, Cho SH, Yun JH, Chae HJ, Ahn YK, *et al.* Effect of green tea consumption on endothelial function and circulating endothelial progenitor cells in chronic smokers. *Circ J* 2006; 70: 1052-7.
- [49] Loke WM, Hodgson JM, Proudfoot JM, McKinley AJ, Puddey IB, Croft KD. Pure dietary flavonoids quercetin and (-)-epicatechin augment nitric oxide products and reduce endothelin-1 acutely in healthy men. *Am J Clin Nutr* 2008; 88: 1018-25.
- [50] Oyama J, Maeda T, Kouzuma K, Ochiai R, Tokimitsu I, Higuchi Y, *et al.* Green tea catechins improve human forearm endothelial dysfunction and have antiatherosclerotic effects in smokers. *Circ J* 2010; 74: 578-88.
- [51] Hodgson JM, Puddey IB, Burke V, Croft KD. Is reversal of endothelial dysfunction by tea related to flavonoid metabolism? *Br J Nutr* 2006; 95: 14-7.
- [52] Duffy SJ, Keaney JF, Jr., Holbrook M, Gokce N, Swerdloff PL, Frei B, *et al.* Short- and long-term black tea consumption reverses endothelial dysfunction in patients with coronary artery disease. *Circulation* 2001; 104: 151-6.
- [53] Hodgson JM. Effects of tea and tea flavonoids on endothelial function and blood pressure: a brief review. *Clin Exp Pharmacol Physiol* 2006; 33: 838-41.
- [54] Jain KS, Kathiravan MK, Somani RS, Shishoo CJ. The biology and chemistry of hyperlipidemia. *Bioorg Med Chem* 2007; 15: 4674-99.
- [55] Imai K, Nakachi K. Cross sectional study of effects of drinking green tea on cardiovascular and liver diseases. *BMJ* 1995; 310: 693-6.
- [56] Zheng XX, Xu YL, Li SH, Liu XX, Hui R, Huang XH. Green tea intake lowers fasting serum total and LDL cholesterol in adults: a meta-analysis of 14 randomized controlled trials. *Am J Clin Nutr* 2011; 94: 601-10.
- [57] Hirano-Ohmori R, Takahashi R, Momiyama Y, Taniguchi H, Yonemura A, Tamai S, *et al.* Green tea consumption and serum malondialdehyde-modified LDL concentrations in healthy subjects. *J Am Coll Nutr* 2005; 24: 342-6.
- [58] Gomikawa S, Ishikawa Y, Hayase W, Haratake Y, Hirano N, Matuura H, *et al.* Effect of ground green tea drinking for 2 weeks on the susceptibility of plasma and LDL to the oxidation ex vivo in healthy volunteers. *Kobe J Med Sci* 2008; 54: E62-72.
- [59] Kim A, Chiu A, Barone MK, Avino D, Wang F, Coleman CI, *et al.* Green tea catechins decrease total and low-density lipoprotein cholesterol: A systematic review and meta-analysis. *J Am Diet Assoc*; 111: 1720-9.
- [60] Hsu CH, Tsai TH, Kao YH, Hwang KC, Tseng TY, Chou P. Effect of green tea extract on obese women: A randomized, double-blind, placebo-controlled clinical trial. *Clin Nutr* 2008; 27: 363-70.
- [61] Nagao T, Hase T, Tokimitsu I. A green tea extract high in catechins reduces body fat and cardiovascular risks in humans. *Obesity (Silver Spring)* 2007; 15: 1473-83.

- [62] Wu AH, Spicer D, Stanczyk FZ, Tseng CC, Yang CS, Pike MC. Effect of 2-month controlled green tea intervention on lipoprotein cholesterol, glucose, and hormone levels in healthy postmenopausal women. *Cancer Prev Res (Phila)* 2012; 5: 393-402.
- [63] Inami S, Takano M, Yamamoto M, Murakami D, Tajika K, Yodogawa K, *et al.* Tea catechin consumption reduces circulating oxidized low-density lipoprotein. *Int Heart J* 2007; 48: 725-32.
- [64] Erba D, Riso P, Bordoni A, Foti P, Biagi PL, Testolin G. Effectiveness of moderate green tea consumption on antioxidative status and plasma lipid profile in humans. *J Nutr Biochem* 2005; 16: 144-9.
- [65] Basu A, Sanchez K, Leyva MJ, Wu M, Betts NM, Aston CE, *et al.* Green tea supplementation affects body weight, lipids, and lipid peroxidation in obese subjects with metabolic syndrome. *J Am Coll Nutr* 2010; 29: 31-40.
- [66] Eichenberger P, Colombani PC, Mettler S. Effects of 3-week consumption of green tea extracts on whole-body metabolism during cycling exercise in endurance-trained men. *Int J Vitam Nutr Res* 2009; 79: 24-33.
- [67] Trautwein EA, Du Y, Meynen E, Yan X, Wen Y, Wang H, *et al.* Purified black tea theaflavins and theaflavins/catechin supplements did not affect serum lipids in healthy individuals with mildly to moderately elevated cholesterol concentrations. *Eur J Nutr* 2010; 49: 27-35.
- [68] Stensvold I, Tverdal A, Solvoll K, Foss OP. Tea consumption, relationship to cholesterol, blood pressure, and coronary and total mortality. *Prev Med* 1992; 21: 546-53.
- [69] Wakabayashi K, Kono S, Shinchi K, Honjo S, Todoroki I, Sakurai Y, *et al.* Habitual coffee consumption and blood pressure: A study of self-defense officials in Japan. *Eur J Epidemiol* 1998; 14: 669-73.
- [70] Yang YC, Lu FH, Wu JS, Wu CH, Chang CJ. The protective effect of habitual tea consumption on hypertension. *Arch Intern Med* 2004; 164: 1534-40.
- [71] Hodgson JM, Devine A, Puddey IB, Chan SY, Beilin LJ, Prince RL. Tea intake is inversely related to blood pressure in older women. *J Nutr* 2003; 133: 2883-6.
- [72] Brown AL, Lane J, Coverly J, Stocks J, Jackson S, Stephen A, *et al.* Effects of dietary supplementation with the green tea polyphenol epigallocatechin-3-gallate on insulin resistance and associated metabolic risk factors: randomized controlled trial. *Br J Nutr* 2009; 101: 886-94.
- [73] Brown AL, Lane J, Holyoak C, Nicol B, Mayes AE, Dadd T. Health effects of green tea catechins in overweight and obese men: A randomised controlled cross-over trial. *Br J Nutr* 2011; 106: 1880-9.
- [74] Potenza MA, Marasciulo FL, Tarquinio M, Tiravanti E, Colantuono G, Federici A, *et al.* EGCG, a green tea polyphenol, improves endothelial function and insulin sensitivity, reduces blood pressure, and protects against myocardial I/R injury in SHR. *Am J Physiol Endocrinol Metab* 2007; 292: E1378-87.
- [75] Sanae F, Miyaichi Y, Kizu H, Hayashi H. Effects of catechins on vascular tone in rat thoracic aorta with endothelium. *Life Sci* 2002; 71: 2553-62.
- [76] Lim DY, Lee ES, Park HG, Kim BC, Hong SP, Lee EB. Comparison of green tea extract and epigallocatechin gallate on blood pressure and contractile responses of vascular smooth muscle of rats. *Arch Pharm Res* 2003; 26: 214-23.
- [77] Ihm SH, Lee JO, Kim SJ, Seung KB, Schini-Kerth VB, Chang K, *et al.* Catechin prevents endothelial dysfunction in the prediabetic stage of OLETF rats by reducing vascular NADPH oxidase activity and expression. *Atherosclerosis* 2009; 206: 47-53.
- [78] Igarashi K, Honma K, Yoshinari O, Nanjo F, Hara Y. Effects of dietary catechins on glucose tolerance, blood pressure and oxidative status in Goto-Kakizaki rats. *J Nutr Sci Vitaminol (Tokyo)* 2007; 53: 496-500.
- [79] Antonello M, Montemurro D, Bolognesi M, Di Pascoli M, Piva A, Grego F, *et al.* Prevention of hypertension, cardiovascular damage and endothelial dysfunction with green tea extracts. *Am J Hypertens* 2007; 20: 1321-8.
- [80] Papparella I, Ceolotto G, Montemurro D, Antonello M, Garbisa S, Rossi G, *et al.* Green tea attenuates angiotensin II-induced cardiac hypertrophy in rats by modulating reactive oxygen species production and the Src/epidermal growth factor receptor/Akt signaling pathway. *J Nutr* 2008; 138: 1596-601.
- [81] Verschuren L, Wielinga PY, van Duyvenvoorde W, Tijani S, Toet K, van Ommen B, *et al.* A dietary mixture containing fish oil, resveratrol, lycopene, catechins, and vitamins E and C reduces atherosclerosis in transgenic mice. *J Nutr* 2011; 141: 863-9.
- [82] Miura Y, Chiba T, Tomita I, Koizumi H, Miura S, Umegaki K, *et al.* Tea catechins prevent the development of atherosclerosis in apolipoprotein E-deficient mice. *J Nutr* 2001; 131: 27-32.
- [83] Chyu KY, Babbidge SM, Zhao X, Dandillaya R, Rietveld AG, Yano J, *et al.* Differential effects of green tea-derived catechin on developing versus established atherosclerosis in apolipoprotein E-null mice. *Circulation* 2004; 109: 2448-53.
- [84] Hirase T, Node K. Endothelial dysfunction as a cellular mechanism for vascular failure. *Am J Physiol Heart Circ Physiol* 2012; 302: H499-505.
- [85] Massion PB, Feron O, Dessy C, Balligand JL. Nitric oxide and cardiac function: ten years after, and continuing. *Circ Res* 2003; 93: 388-98.
- [86] Deanfield JE, Halcox JP, Rabelink TJ. Endothelial function and dysfunction: testing and clinical relevance. *Circulation* 2007; 115: 1285-95.
- [87] Vita JA. Endothelial function. *Circulation* 2011; 124: e906-12.
- [88] Jochmann N, Lorenz M, Krosigk A, Martus P, Bohm V, Baumann G, *et al.* The efficacy of black tea in ameliorating endothelial function is equivalent to that of green tea. *Br J Nutr* 2008; 99: 863-8.
- [89] Lorenz M, Wessler S, Follmann E, Michaelis W, Dusterhoft T, Baumann G, *et al.* A constituent of green tea, epigallocatechin-3-gallate, activates endothelial nitric oxide synthase by a phosphatidylinositol-3-OH-kinase-, cAMP-dependent protein kinase-, and Akt-dependent pathway and leads to endothelial-dependent vasorelaxation. *J Biol Chem* 2004; 279: 6190-5.
- [90] Benito S, Lopez D, Saiz MP, Buxaderas S, Sanchez J, Puig-Parellada P, *et al.* A flavonoid-rich diet increases nitric oxide production in rat aorta. *Br J Pharmacol* 2002; 135: 910-6.
- [91] Persson IA, Josefsson M, Persson K, Andersson RG. Tea flavanols inhibit angiotensin-converting enzyme activity and increase nitric oxide production in human endothelial cells. *J Pharm Pharmacol* 2006; 58: 1139-44.
- [92] Kim JA, Formoso G, Li Y, Potenza MA, Marasciulo FL, Montagnani M, *et al.* Epigallocatechin gallate, a green tea polyphenol, mediates NO-dependent vasodilation using signaling pathways in vascular endothelium requiring reactive oxygen species and Fyn. *J Biol Chem* 2007; 282: 13736-45.
- [93] Bourque SL, Davidge ST, Adams MA. The interaction between endothelin-1 and nitric oxide in the vasculature: new perspectives. *Am J Physiol Regul Integr Comp Physiol* 2011; 300: R1288-95.
- [94] Yanagisawa M, Kurihara H, Kimura S, Tomobe Y, Kobayashi M, Mitsui Y, *et al.* A novel potent vasoconstrictor peptide produced by vascular endothelial cells. *Nature* 1988; 332: 411-5.
- [95] Gossel M, Lerman A. Endothelin: beyond a vasoconstrictor. *Circulation* 2006; 113: 1156-8.
- [96] de Andrade CR, Leite PF, Montezano AC, Casolari DA, Yogi A, Tostes RC, *et al.* Increased endothelin-1 reactivity and endothelial dysfunction in carotid arteries from rats with hyperhomocysteinemia. *Br J Pharmacol* 2009; 157: 568-80.
- [97] Zhou J, Zhu Y, Cheng M, Dinesh D, Thorne T, Poh KK, *et al.* Regulation of vascular contractility and blood pressure by the E2F2 transcription factor. *Circulation* 2009; 120: 1213-21.
- [98] Thijssen DH, Rongen GA, van Dijk A, Smits P, Hopman MT. Enhanced endothelin-1-mediated leg vascular tone in healthy older subjects. *J Appl Physiol* 2007; 103: 852-7.
- [99] Reiter CE, Kim JA, Quon MJ. Green tea polyphenol epigallocatechin gallate reduces endothelin-1 expression and secretion in vascular endothelial cells: Roles for AMP-activated protein kinase, Akt, and FOXO1. *Endocrinology* 2010; 151: 103-14.
- [100] Tuttolomondo A, Di Raimondo D, Pecoraro R, Arnao V, Pinto A, Licata G. Atherosclerosis as an Inflammatory Disease. *Curr Pharm Des* 2012; [Epub ahead of print].
- [101] Kevil CG, Patel RP, Bullard DC. Essential role of ICAM-1 in mediating monocyte adhesion to aortic endothelial cells. *Am J Physiol Cell Physiol* 2001; 281: C1442-7.
- [102] Gerszten RE, Garcia-Zepeda EA, Lim YC, Yoshida M, Ding HA, Gimbrone MA, Jr., *et al.* MCP-1 and IL-8 trigger firm adhesion of monocytes to vascular endothelium under flow conditions. *Nature* 1999; 398: 718-23.
- [103] Ramesh E, Geraldine P, Thomas PA. Regulatory effect of epigallocatechin gallate on the expression of C-reactive protein and other inflammatory markers in an experimental model of atherosclerosis. *Chem Biol Interact* 2010; 183: 125-32.

- [104] Wang CJ, Liu JT, Guo F. (-)-epigallocatechin gallate inhibits endothelin-1-induced C-reactive protein production in vascular smooth muscle cells. *Basic Clin Pharmacol Toxicol* 2010; 107: 669-75.
- [105] da Silva CG, Specht A, Wegiel B, Ferran C, Kaczmarek E. Mechanism of purinergic activation of endothelial nitric oxide synthase in endothelial cells. *Circulation* 2009; 119: 871-9.
- [106] Ahn HY, Kim CH, Ha TS. Epigallocatechin-3-gallate Regulates NADPH Oxidase Expression in Human Umbilical Vein Endothelial Cells. *Korean J Physiol Pharmacol* 2010; 14: 325-9.
- [107] Ludwig A, Lorenz M, Grimbo N, Steidle F, Meiners S, Bartsch C, *et al.* The tea flavonoid epigallocatechin-3-gallate reduces cytokine-induced VCAM-1 expression and monocyte adhesion to endothelial cells. *Biochem Biophys Res Commun* 2004; 316: 659-65.
- [108] Suzuki J, Ogawa M, Izawa A, Sagesaka YM, Isobe M. Dietary consumption of green tea catechins attenuate hyperlipidaemia-induced atherosclerosis and systemic organ damage in mice. *Acta Cardiol* 2005; 60: 271-6.
- [109] Blackwell TS, Christman JW. The role of nuclear factor-kappa B in cytokine gene regulation. *Am J Respir Cell Mol Biol* 1997; 17: 3-9.
- [110] Ghosh S, May MJ, Kopp EB. NF-kappa B and Rel proteins: Evolutionarily conserved mediators of immune responses. *Annu Rev Immunol* 1998; 16: 225-60.
- [111] Hong MH, Kim MH, Chang HJ, Kim NH, Shin BA, Ahn BW, *et al.* (-)-Epigallocatechin-3-gallate inhibits monocyte chemotactic protein-1 expression in endothelial cells via blocking NF-kappaB signaling. *Life Sci* 2007; 80: 1957-65.
- [112] Ahn HY, Xu Y, Davidge ST. Epigallocatechin-3-O-gallate inhibits TNFalpha-induced monocyte chemotactic protein-1 production from vascular endothelial cells. *Life Sci* 2008; 82: 964-8.
- [113] Pullikotil P, Chen H, Muniyappa R, Greenberg CC, Yang S, Reiter CE, *et al.* Epigallocatechin gallate induces expression of heme oxygenase-1 in endothelial cells via p38 MAPK and Nrf-2 that suppresses proinflammatory actions of TNF-alpha. *J Nutr Biochem* 2011: [Epub ahead of print].
- [114] Lambert JD, Elias RJ. The antioxidant and pro-oxidant activities of green tea polyphenols: a role in cancer prevention. *Arch Biochem Biophys* 2010; 501: 65-72.
- [115] Ying CJ, Xu JW, Ikeda K, Takahashi K, Nara Y, Yamori Y. Tea polyphenols regulate nicotinamide adenine dinucleotide phosphate oxidase subunit expression and ameliorate angiotensin II-induced hyperpermeability in endothelial cells. *Hypertens Res* 2003; 26: 823-8.
- [116] Khan SG, Katiyar SK, Agarwal R, Mukhtar H. Enhancement of antioxidant and phase II enzymes by oral feeding of green tea polyphenols in drinking water to SKH-1 hairless mice: possible role in cancer chemoprevention. *Cancer Res* 1992; 52: 4050-2.
- [117] Paquay JB, Haenen GR, Stender G, Wiseman SA, Tijburg LB, Bast A. Protection against nitric oxide toxicity by tea. *J Agric Food Chem* 2000; 48: 5768-72.
- [118] Abo A, Pick E, Hall A, Totty N, Teahan CG, Segal AW. Activation of the NADPH oxidase involves the small GTP-binding protein p21rac1. *Nature* 1991; 353: 668-70.
- [119] Sakamoto N, Ishibashi T, Sugimoto K, Sawamura T, Sakamoto T, Inoue N, *et al.* Role of LOX-1 in monocyte adhesion-triggered redox, Akt/eNOS and Ca<sup>2+</sup> signaling pathways in endothelial cells. *J Cell Physiol* 2009; 220: 706-15.
- [120] Schewe T, Steffen Y, Sies H. How do dietary flavanols improve vascular function? A position paper. *Arch Biochem Biophys* 2008; 476: 102-6.
- [121] Mertens A, Holvoet P. Oxidized LDL and HDL: Antagonists in atherothrombosis. *Faseb J* 2001; 15: 2073-84.
- [122] Tijburg LB, Wiseman SA, Meijer GW, Weststrate JA. Effects of green tea, black tea and dietary lipophilic antioxidants on LDL oxidizability and atherosclerosis in hypercholesterolaemic rabbits. *Atherosclerosis* 1997; 135: 37-47.
- [123] Yamanaka N, Oda O, Nagao S. Green tea catechins such as (-)-epicatechin and (-)-epigallocatechin accelerate Cu<sup>2+</sup>-induced low density lipoprotein oxidation in propagation phase. *FEBS Lett* 1997; 401: 230-4.
- [124] Ishikawa T, Suzukawa M, Ito T, Yoshida H, Ayaori M, Nishiwaki M, *et al.* Effect of tea flavonoid supplementation on the susceptibility of low-density lipoprotein to oxidative modification. *Am J Clin Nutr* 1997; 66: 261-6.
- [125] Ohmori R, Iwamoto T, Tago M, Takeo T, Unno T, Itakura H, *et al.* Antioxidant activity of various teas against free radicals and LDL oxidation. *Lipids* 2005; 40: 849-53.
- [126] Chen XP, Xun KL, Wu Q, Zhang TT, Shi JS, Du GH. Oxidized low density lipoprotein receptor-1 mediates oxidized low density lipoprotein-induced apoptosis in human umbilical vein endothelial cells: role of reactive oxygen species. *Vascul Pharmacol* 2007; 47: 1-9.
- [127] Ou HC, Song TY, Yeh YC, Huang CY, Yang SF, Chiu TH, *et al.* EGCG protects against oxidized LDL-induced endothelial dysfunction by inhibiting LOX-1-mediated signaling. *J Appl Physiol* 2010; 108: 1745-56.
- [128] Sheng R, Gu ZL, Xie ML, Zhou WX, Guo CY. Epigallocatechin gallate protects H9c2 cardiomyoblasts against hydrogen dioxides-induced apoptosis and telomere attrition. *Eur J Pharmacol* 2010; 641: 199-206.
- [129] Yang J, Han Y, Sun H, Chen C, He D, Guo J, *et al.* (-)-Epigallocatechin gallate suppresses proliferation of vascular smooth muscle cells induced by high glucose by inhibition of PKC and ERK1/2 signalings. *J Agric Food Chem* 2011; 59: 11483-90.
- [130] Won SM, Park YH, Kim HJ, Park KM, Lee WJ. Catechins inhibit angiotensin II-induced vascular smooth muscle cell proliferation via mitogen-activated protein kinase pathway. *Exp Mol Med* 2006; 38: 525-34.
- [131] Han DW, Jung DY, Park JC, Cho HH, Hyon SH, Han DK. Underlying mechanism for suppression of vascular smooth muscle cells by green tea polyphenol EGCG released from biodegradable polymers for stent application. *J Biomed Mater Res A* 2010; 95: 424-33.
- [132] Chan CM, Huang JH, Chiang HS, Wu WB, Lin HH, Hong JY, *et al.* Effects of (-)-epigallocatechin gallate on RPE cell migration and adhesion. *Mol Vis* 2010; 16: 586-95.
- [133] Park JS, Kim MH, Chang HJ, Kim KM, Kim SM, Shin BA, *et al.* Epigallocatechin-3-gallate inhibits the PDGF-induced VEGF expression in human vascular smooth muscle cells via blocking PDGF receptor and Erk-1/2. *Int J Oncol* 2006; 29: 1247-52.
- [134] Weber AA, Neuhaus T, Skach RA, Hescheler J, Ahn HY, Schror K, *et al.* Mechanisms of the inhibitory effects of epigallocatechin-3 gallate on platelet-derived growth factor-BB-induced cell signaling and mitogenesis. *Faseb J* 2004; 18: 128-30.
- [135] Newby AC. Matrix metalloproteinase inhibition therapy for vascular diseases. *Vascul Pharmacol* 2012; 56: 232-44. .
- [136] El Bedoui J, Oak MH, Anglard P, Schini-Kerth VB. Catechins prevent vascular smooth muscle cell invasion by inhibiting MT1-MMP activity and MMP-2 expression. *Cardiovasc Res* 2005; 67: 317-25.
- [137] Maeda K, Kuzuya M, Cheng XW, Asai T, Kanda S, Tamaya-Mori N, *et al.* Green tea catechins inhibit the cultured smooth muscle cell invasion through the basement barrier. *Atherosclerosis* 2003; 166: 23-30.
- [138] Kim CH, Moon SK. Epigallocatechin-3-gallate causes the p21/WAF1-mediated G(1)-phase arrest of cell cycle and inhibits matrix metalloproteinase-9 expression in TNF-alpha-induced vascular smooth muscle cells. *Arch Biochem Biophys* 2005; 435: 264-72.
- [139] Gouni-Berthold I, Sachinidis A. Molecular mechanisms explaining the preventive effects of catechins on the development of proliferative diseases. *Curr Pharm Des* 2004; 10: 1261-71.
- [140] Vorchheimer DA, Becker R. Platelets in atherothrombosis. *Mayo Clin Proc* 2006; 81: 59-68.
- [141] Kang WS, Lim IH, Yuk DY, Chung KH, Park JB, Yoo HS, *et al.* Antithrombotic activities of green tea catechins and (-)-epigallocatechin gallate. *Thromb Res* 1999; 96: 229-37.
- [142] Kang WS, Chung KH, Chung JH, Lee JY, Park JB, Zhang YH, *et al.* Antiplatelet activity of green tea catechins is mediated by inhibition of cytoplasmic calcium increase. *J Cardiovasc Pharmacol* 2001; 38: 875-84.
- [143] Son DJ, Cho MR, Jin YR, Kim SY, Park YH, Lee SH, *et al.* Antiplatelet effect of green tea catechins: a possible mechanism through arachidonic acid pathway. *Prostaglandins Leukot Essent Fatty Acids* 2004; 71: 25-31.
- [144] Jin YR, Im JH, Park ES, Cho MR, Han XH, Lee JJ, *et al.* Antiplatelet effect of epigallocatechin gallate is mediated by the inhibition of PLCgamma2 phosphorylation, elevation of PGD2 production, and maintaining calcium-ATPase activity. *J Cardiovasc Pharmacol* 2008; 51: 45-54.
- [145] Matsumura K, Takayama H, Bae JY, Kurihara M, Tsutsumi S, Hyon SH. Preservation of platelets by adding epigallocatechin-3-o-gallate to platelet concentrates. *Cell Transplant* 2009; 18: 521-8.

- [146] Lill G, Voit S, Schror K, Weber AA. Complex effects of different green tea catechins on human platelets. *FEBS Lett* 2003; 546: 265-70.
- [147] Yang JA, Choi JH, Rhee SJ. Effects of green tea catechin on phospholipase A2 activity and antithrombus in streptozotocin diabetic rats. *J Nutr Sci Vitaminol (Tokyo)* 1999; 45: 337-46.
- [148] Vezza R, Mezzasoma AM, Venditti G, Greslele P. Prostaglandin endoperoxidase and thromboxane A2 activate the same receptor isoforms in human platelets. *Thromb Haemostasis* 2002; 87: 114-21.
- [149] Sugatani J, Fukazawa N, Ujihara K, Yoshinari K, Abe I, Noguchi H, *et al.* Tea polyphenols inhibit acetyl-CoA:1-alkyl-sn-glycerol-3-phosphocholine acetyltransferase (a key enzyme in platelet-activating factor biosynthesis) and platelet-activating factor-induced platelet aggregation. *Int Arch Allergy Immunol* 2004; 134: 17-28.
- [150] Senthil KV, Arulmathi K, Sundarapandian R, Kalaiselvi P. Attenuation of the inflammatory changes and lipid anomalies by epigallocatechin-3-gallate in hypercholesterolemic diet fed aged rats. *Exp Gerontol* 2009; 44: 745-51.
- [151] Anandh Babu PV, Sabitha KE, Shyamaladevi CS. Green tea extract impedes dyslipidaemia and development of cardiac dysfunction in streptozotocin-diabetic rats. *Clin Exp Pharmacol Physiol* 2006; 33: 1184-9.
- [152] Babu PV, Sabitha KE, Shyamaladevi CS. Green tea impedes dyslipidemia, lipid peroxidation, protein glycation and ameliorates  $Ca^{2+}$ -ATPase and  $Na^{+}/K^{+}$ -ATPase activity in the heart of streptozotocin-diabetic rats. *Chem Biol Interact* 2006; 162: 157-64.
- [153] Yang TT, Koo MW. Hypocholesterolemic effects of Chinese tea. *Pharmacol Res* 1997; 35: 505-12.
- [154] Roghani M, Baluchnejadmojarad T. Hypoglycemic and hypolipidemic effect and antioxidant activity of chronic epigallocatechin-gallate in streptozotocin-diabetic rats. *Pathophysiology* 2010; 17: 55-9.
- [155] Bursill CA, Abbey M, Roach PD. A green tea extract lowers plasma cholesterol by inhibiting cholesterol synthesis and upregulating the LDL receptor in the cholesterol-fed rabbit. *Atherosclerosis* 2007; 193: 86-93.
- [156] Koo SI, Noh SK. Green tea as inhibitor of the intestinal absorption of lipids: potential mechanism for its lipid-lowering effect. *J Nutr Biochem* 2007; 18: 179-83.
- [157] Wang S, Noh SK, Koo SI. Green tea catechins inhibit pancreatic phospholipase A(2) and intestinal absorption of lipids in ovariectomized rats. *J Nutr Biochem* 2006; 17: 492-8.
- [158] Raederstorff DG, Schlachter MF, Elste V, Weber P. Effect of EGCG on lipid absorption and plasma lipid levels in rats. *J Nutr Biochem* 2003; 14: 326-32.
- [159] Cuccioloni M, Mozzicafreddo M, Spina M, Tran CN, Falconi M, Eleuteri AM, *et al.* Epigallocatechin-3-gallate potently inhibits the *in vitro* activity of hydroxy-3-methyl-glutaryl-CoA reductase. *J Lipid Res* 2011; 52: 897-907.
- [160] Abe I, Seki T, Umehara K, Miyase T, Noguchi H, Sakakibara J, *et al.* Green tea polyphenols: Novel and potent inhibitors of squalene epoxidase. *Biochem Biophys Res Commun* 2000; 268: 767-71.
- [161] Ikeda I, Imasato Y, Sasaki E, Nakayama M, Nagao H, Takeo T, *et al.* Tea catechins decrease micellar solubility and intestinal absorption of cholesterol in rats. *Biochim Biophys Acta* 1992; 1127: 141-6.
- [162] Shishikura Y, Khokhar S, Murray BS. Effects of tea polyphenols on emulsification of olive oil in a small intestine model system. *J Agric Food Chem* 2006; 54: 1906-13.
- [163] Chan PT, Fong WP, Cheung YL, Huang Y, Ho WK, Chen ZY. Jasmine green tea epicatechins are hypolipidemic in hamsters (*Mesocricetus auratus*) fed a high fat diet. *J Nutr* 1999; 129: 1094-101.
- [164] Leslie EM, Mao Q, Oleschuk CJ, Deeley RG, Cole SP. Modulation of multidrug resistance protein 1 (MRP1/ABCC1) transport and at-pase activities by interaction with dietary flavonoids. *Mol Pharmacol* 2001; 59: 1171-80.
- [165] Lee MS, Park JY, Freaque H, Kwun IS, Kim Y. Green tea catechin enhances cholesterol 7 $\alpha$ -hydroxylase gene expression in HepG2 cells. *Br J Nutr* 2008; 99: 1182-5.
- [166] Goto T, Saito Y, Morikawa K, Kanamaru Y, Nagaoka S. Epigallocatechin gallate changes mRNA expression level of genes involved in cholesterol metabolism in hepatocytes. *Br J Nutr* 2012; 107: 769-73.
- [167] Villarreal F, Yamazaki, K.G., Taub, P.R., Maisel, A. Use of epicatechin and derivatives and salts thereof for cardiac protection of ischemic myocardium and ameliorate adverse cardiac remodeling. US20110021466 A1 (2011).
- [168] Bukowski, JF, Walters, J. Tea-derived compositions and methods of using same for cardiovascular health. US20080081066A1 (2008).
- [169] Raederstorff, D, Weber, P., Wolfram, S. Nutraceutical compositions comprising epigallocatechin gallate and raspberry ketone. US20090221694A1 (2009).
- [170] Unno, T., Kakuda, T., Miyazawa, T., Nakagawa, K. Method for improving absorbability of epigallocatechin gallate, foods, drinks and food/drink materials using the same and method for producing the same. US20090047408A1 (2009).
- [171] Collin, P.D., Tilson, III, M.D. Methods for treating or preventing a vascular disease. US20080161009A1 (2008).
- [172] Sone T, Kuriyama S, Nakaya N, Hozawa A, Shimazu T, Nomura K, *et al.* Randomized controlled trial for an effect of catechin-enriched green tea consumption on adiponectin and cardiovascular disease risk factors. *Food Nutr Res* 2011; 55.
- [173] Hirano R, Momiyama Y, Takahashi R, Taniguchi H, Kondo K, Nakamura H, *et al.* Comparison of green tea intake in Japanese patients with and without angiographic coronary artery disease. *Am J Cardiol* 2002; 90: 1150-3.
- [174] Yang CS, Chen L, Lee MJ, Balentine D, Kuo MC, Schantz SP. Blood and urine levels of tea catechins after ingestion of different amounts of green tea by human volunteers. *Cancer Epidemiol Biomarkers Prev* 1998; 7: 351-4.
- [175] Lee MJ, Wang ZY, Li H, Chen L, Sun Y, Gobbo S, *et al.* Analysis of plasma and urinary tea polyphenols in human subjects. *Cancer Epidemiol Biomarkers Prev* 1995; 4: 393-9.
- [176] Auger C, Mullen W, Hara Y, Crozier A. Bioavailability of polyphenon E flavan-3-ols in humans with an ileostomy. *J Nutr* 2008; 138: 1535S-42S.
- [177] Chow HH, Hakim IA, Vining DR, Crowell JA, Ranger-Moore J, Chew WM, *et al.* Effects of dosing condition on the oral bioavailability of green tea catechins after single-dose administration of Polyphenon E in healthy individuals. *Clin Cancer Res* 2005; 11: 4627-33.
- [178] Barras A, Mezzetti A, Richard A, Lazzaroni S, Roux S, Melnyk P, *et al.* Formulation and characterization of polyphenol-loaded lipid nanocapsules. *Int J Pharm* 2009; 379: 270-7.
- [179] Siddiqui IA, Adhami VM, Bharali DJ, Hafeez BB, Asim M, Khwaja SI, *et al.* Introducing nanochemoprevention as a novel approach for cancer control: proof of principle with green tea polyphenol epigallocatechin-3-gallate. *Cancer Res* 2009; 69: 1712-6.
- [180] Smith A, Giunta B, Bickford PC, Fountain M, Tan J, Shytle RD. Nanolipidic particles improve the bioavailability and alpha-secretase inducing ability of epigallocatechin-3-gallate (EGCG) for the treatment of Alzheimer's disease. *Int J Pharm* 2010; 389: 207-12.
- [181] Hsu, S. Modified green tea polyphenol formulations. US2012076872A1 (2012).
- [182] Lambelet, P., Bortlik, K., Sabatier, M., Crespy, V., Williamson, G. Green tea extracts of improved bioavailability. EP2434911A1 (2012).
- [183] Townsend PA, Scarabelli TM, Pasini E, Gitti G, Menegazzi M, Suzuki H, *et al.* Epigallocatechin-3-gallate inhibits STAT-1 activation and protects cardiac myocytes from ischemia/reperfusion-induced apoptosis. *Faseb J* 2004; 18: 1621-3.
- [184] Williamson G, Manach C. Bioavailability and bioefficacy of polyphenols in humans. II. Review of 93 intervention studies. *Am J Clin Nutr* 2005; 81: 243S-55S.
- [185] Chow HH, Cai Y, Hakim IA, Crowell JA, Shahi F, Brooks CA, *et al.* Pharmacokinetics and safety of green tea polyphenols after multiple-dose administration of epigallocatechin gallate and polyphenon E in healthy individuals. *Clin Cancer Res* 2003; 9: 3312-9.